BIMM 101 - Recombinant DNA Techniques, Winter 2022

Welcome!

Time and Place:

<u>Lecture</u>: Tues/Thurs 8:00 – 9:20a, HSS 1330 or Zoom (will be recorded)

<u>Lab</u>: Tues/Thurs 10:00a – 1:50p, York 2310/2332 or Zoom (D01/D02)

Wed/Fri 9:00 – 12:50p, York 2310/2332 or Zoom (D03/D04)

Instructor:

Emily Grossman, PhD H&SS 1145H

egrossma@ucsd.edu

In person Office Hours: Mondays 1 – 2p, H&SS 1145L Zoom Office Hours: Thursdays 3 – 4p, Zoom link on Canvas

Instructional Assistants:

Irene Choi <u>ivchoi@ucsd.edu</u>
Yuxi Wei <u>yuwei@ucsd.edu</u>
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Learning goals:

- Learn the theory behind molecular techniques, and the applications of the methodologies in biological research
- Become proficient at basic molecular biology techniques
- Learn the importance of proper controls in designing experiments and interpreting results
- Improve lab math skills and ability to graph data correctly
- Learn to make logical conclusions from experimental data
- Become familiar with bioinformatics databases and applications
- Learn to find, read, and evaluate primary literature
- Become aware of the implications of the technology for society

Learning in this Course:

This course is designed to be an environment for everyone to learn and construct a shared understanding of the material. **Active participation** by engaging with the lecture material, asking and answering questions (e.g. on the discussion board), and contributing to breakout sessions during lab time is expected. Being able to communicate understanding, and confusion, is critical to success in any discipline, and is very useful for learning. To encourage collaboration, lab discussions will be done in groups, and grades will not be assigned on a curve. Being proactive to ask questions during office hours and on the discussion board will be critical for success, especially given the online nature of the course.

Instead of memorization, we will focus on developing an understanding of fundamental concepts as they apply to different examples. Therefore, quizzes will include questions that are based on solving problems in new contexts.

Resources

1. Support for student writers:

Writing + Critical Expression Hub (part of the Teaching + Learning Commons)

There are undergraduate writing mentors on staff who are in Biological Sciences degree programs, so they have training not only in working with student writers, but also familiarity with science reasoning and science writing. Please don't hesitate to meet with a writing mentor for help—every writer can benefit from these conversations. Students can make appointments via https://ucsd.mywconline.com

2. Tutoring

OASIS: Office of Academic Support and Instructional Services

From the OASIS website (https://students.ucsd.edu/sponsor/oasis/): We are the learning center at UC San Diego and provide most of the free tutoring on campus in a collaborative, supportive environment. All UC San Diego students are eligible to receive OASIS services. Each year, OASIS serves 3,000 students in language, math, science, study skills, and writing as well as peer counseling and peer mentoring. They are located on the third floor of Center Hall, (858) 534-3760 (phone), oasis@ucsd.edu (email)

Required Materials

- 1. Labcoat must go to knees (available at bookstore)
- 2. UV blocking safety glasses (available at bookstore)
- 3. BIMM 101 Lab manual (available at bookstore)
- 4. Notebook (with or without carbon copies is fine)
- 5. Fine point Sharpie for labeling dark color (blue/black) is best
- 6. Long pants and closed-toed shoes are always required in lab (entire legs and feet covered, including ankles)

Lab Safety Training: Enrolled and waitlisted students MUST successfully complete the Biology Lab Safety Training and Assessment before the first lab session: https://biolabclass-safetyquiz.ucsd.edu/introduction. Please note that courses offered by other departments (Chemistry, for example) may have additional safety training requirements.

Attendance: Enrolled and waitlisted students MUST attend the first lab session. Additional details: http://biology.ucsd.edu/go/ug-labs. Remember that lab attendance is required – if you miss more than two in person labs, you will be asked to drop the course, or will receive an F for the course. If you are ill, you must leave a message with me, not your IA. Please be on time for lab; the IAs go over the experiments at the beginning of lab. If you are habitually late to lab, you will lose 5% from your final grade.

Add/Drop Deadlines are different for lab courses than lecture courses. Students who drop a Biology lab class after the end of the second class meeting will be assigned a "W". Additional details: http://biology.ucsd.edu/go/ug-labs.

Accessibility

Any student with a disability is welcome to contact me in order to work out reasonable accommodations to support their success in this course. Students requesting accommodations for this course due to a disability must work through the Office for Students with Disabilities (OSD). Instructors will receive Authorization for Accommodations Letters from the OSD online portal. Whenever possible, we will use universal designs that are inclusive. If you have feedback on how to make the class more accessible and inclusive, please get in touch!

If you received accommodations during Fall 2021 and would like to request the same accommodations for Winter 2022, please email your OSD specialist using the subject line "Requesting Winter Accommodations." Your Specialist will review your file and let you know if you need to provide updated documentation for the Winter Quarter before accommodations can be determined.

Students Who Have Not Received Accommodations Through The OSD Previously This includes those with temporary limitations (concussions, broken bones, etc.)

All intake appointments will take place over the phone/Zoom. Call the OSD at 858.534.4382 to schedule an appointment. If campus is closed, send an email to osd@ucsd.edu and put "Request for Intake Appointment" in the subject line. Indicate dates and times during the subsequent two weeks that you are available for a 60 minute phone appointment, and you will be contacted via phone or email. If you are deaf or hard of hearing and would prefer to have your intake conducted via email or through Zoom with captions, please let us know.

Before your appointment, complete the intake and consent forms which may be found on the OSD website: osd.ucsd.edu/students/forms.html#Student-Forms and email them back to osd@ucsd.edu. These will be forwarded to your Disability Specialist before your appointment.

Contact the OSD for further information:

858.534.4382 (phone) osd@ucsd.edu (email) http://disabilities.ucsd.edu (website)

Inclusion

It is our goal to create a learning environment that supports diversity of thought, perspective, experience, and identities. We encourage all of you to participate in discussion and contribute to the field from your perspective. If you have feedback on how to make the class more inclusive, please get in touch!

Office of Equity, Diversity, and Inclusion:

858.822.3542 | diversity@ucsd.edu | https://diversity.ucsd.edu/ https://students.ucsd.edu/student-life/diversity/index.html https://regents.universityofcalifornia.edu/governance/policies/4400.html

A Culture of Respect: The Office for the Prevention of Harassment & Discrimination (OPHD) provides assistance to students, faculty, and staff regarding reports of bias, harassment, and discrimination. Students have the right to an educational environment that is free from harassment and discrimination.

Students have options for reporting incidents of sexual violence and sexual harassment. Sexual violence includes sexual assault, dating violence, domestic violence, and stalking. Information about reporting options may be obtained at OPHD at (858) 534-8298, ophd@ucsd.edu, or http://ophd.ucsd.edu. Students may receive confidential assistance at the Sexual Assault Resource Center at (858) 534-5793, sarc@ucsd.edu, or http://care.ucsd.edu, or through Counseling and Psychological Services (CAPS) at (858) 534-3755 or http://caps.ucsd.edu

Students may feel more comfortable discussing their particular concern with a trusted employee. This may be a UCSD student affairs staff member, a department Chair, a faculty member or other University official. These individuals have an obligation to report incidents of sexual violence and sexual harassment to OPHD. This does not necessarily mean that a formal complaint will be filed.

If you find yourself in an uncomfortable situation, ask for help. UCSD is committed to upholding policies regarding nondiscrimination, sexual violence and sexual harassment.

Class Web Site:

The class web site is on Canvas (https://coursefinder.ucsd.edu) All class notices, the syllabus, and other important information will be posted here. Please check the web site regularly for updates, since this will be the main form of distribution of information to the class. My lecture notes will be posted to the site.

Technical Support:

For help with accounts, network, and technical issues:

https://acms.ucsd.edu/contact/index.html

For help connecting to electronic library resources such as eReserves and e-journals: https://library.ucsd.edu/computing-and-technology/connect-from-off-campus/

Course Requirements and Grading:

The class will be out of 500 points total, with the following components and each of their respective percentages of the total.

Grading Component	Points	Percentage
Post-lecture quizzes (over 85% correct earns a full score)	25	5.0%
Zoom Lab Discussion (attendance and participation at 85% of zoom sessions earns full score)	25	5.00/
Lab Notebooks	25 100	5.0% 20.0%
Problem Sets (8)	175	35.0%
Lab report #1 - Oligo design	40	8.0%
Lab report #2 - CRISPR Writeup	110	22.0%
Technique Presentation and Writeup	25	5.0%

The following grading scheme will be used. The course is <u>not</u> graded on a curve (i.e. 20% of students getting A, B, C, and such). Thus, the ability to do well in this course is not dependent on others doing poorly.

Total	Grade	80.0 – 81.9%	B-
percentage			
98.0 – 100%	A+	78.0 – 79.9%	C+
92.0 – 97.9%	Α	72.0 – 77.9%	С
90.0 – 91.9%	A-	70.0 – 71.9%	C-
88.0 - 89.9%	B+	60.0 – 69.9%	D
82.0 – 87.9%	В	<59.9%	F

Due to the generous amount of extra credit opportunities, I do not round final scores.

1. Post lecture quizzes, 5%

The quizzes posted on Canvas are meant to reinforce importance concepts covered during lectures. Quizzes are to be completed *prior* to the start of the following lecture. (deadlines will be posted on Canvas). Because mastery is not necessarily expected after watching/attending the lecture, scoring 85% or higher overall will result in full points. If your average score overall on the quizzes falls below 85%, you will earn your average percent (out of 25 points) for this portion of the grade. It is very important to follow-up in office hours or in lab on concepts you were unclear on.

2. Zoom Lab Discussion Participation, 5%

Links to join the video lab sessions will be provided on Canvas. Discussions will be facilitated by the Instructional Assistants and Instructor and are meant to be a time to work collaboratively to analyze data, design experiments, and engage in troubleshooting of results. Participation in at least 85% of the online lab sessions will

result in full points. It is highly recommended that you participate in as many as possible because this is an opportunity to ask questions and get feedback. If circumstances beyond your control interfere with your ability to participate, please get in touch with me so we can devise a plan for you to succeed in the course.

3. Lab notebook (see page 6 in lab manual), 20%:

It is mandatory that you keep a lab notebook, which your IA's will check at the end of every lab for completeness. It should include:

- Purpose: objective of the lab in your own words (why are you doing the experiment?)
- Methods: pages of protocol/procedure and any changes you made to it, relevant charts
- Results: all calculations and data you collect, observations
- Conclusions: summarize and interpret results, labeling & location of samples

We may periodically collect and grade specific assignments.

- **4. Problem Sets: 35%** There will be eight Problem Sets due throughout the quarter. Each Problem set is worth 5% of your final grade. The problem sets will cover the lectures, readings, and lab experiments from the previous week. **Your lowest score will be dropped.** Starting Week 1, Problem Sets will be released on Friday mornings and due to Canvas the following Tuesday at 11:59pm (see calendar on Canvas).
- 5. Lab reports (2): 30% total. You will turn two written reports that are based on data and work from the majority of the quarter. Guidelines will be posted on Canvas and due dates will be on the Canvas calendar. Lab reports must be submitted to Turnitin on Canvas. Although you will be working together to analyze data with your lab group, you must hand in your own assignments, written in your own words. Copying someone else's homework or lab reports (including past quarters!) is cheating (see Academic Integrity statement below).

6. Technique Presentation and Writeup, 5%

At the end of the quarter there will group project presentations. Guidelines for this assignment will be posted on Canvas and due dates will be on the Canvas calendar.

7. Extra credit opportunities (opportunity to earn up to 10pts (2%))

Extra credit problems will be presented throughout the quarter on Problem Sets. In addition, if 85% of the class completes a CAPE evaluation at the end of the quarter for me, everyone will receive 1pt of extra credit as well.

Late policy:

Assignments must be submitted on time to be eligible for full credit. Except in the case of medical or family emergencies, late assignments will be subjected to a 5% deduction per day if submitted within 72 hours after the posted due date. Assignments not submitted within 72 hours of the due date will receive a score of 0.

Regrades:

If a grading error has been made, you should submit a re-grade request to your Instructional Assistant or Dr. Grossman. Students who submit items for re-grading understand that we may re-grade the entire item and the score may go up or down.

Academic Integrity:

We hold the following values (adapted from the International Center for Academic Integrity) as important to academic integrity and for maintaining an inclusive classroom environment. Although we will expect students to work together, all lab reports for the class must be independently written, i.e., **your own work in your own words**. While discussion of data among lab partners is encouraged, each student on their own must complete all text, references, figures, graphs, and tables. If you have questions about the difference between discussing your work with others and unauthorized collaboration, please ask your instructor or IA for clarification. Directly copying material from other sources without putting it in your own words is also plagiarism, even if the source is cited as a reference (including the lab manual for this class! Please put it into your own words!)

	As students, this means you will	As the Instructional team, this means we will
Honesty	 Honestly demonstrate your knowledge and abilities Communicate openly without using deception, including citing sources appropriately 	 Give you honest feedback on your demonstration of knowledge and abilities on assignments and exams Communicate openly and honestly about the expectations and standards of the course through the syllabus and guidelines for course assignments
Responsibilities	 Complete your assignments on time and be fully prepared for class Arrive to lecture and lab on time and be active participants 	 Give you timely feedback on your assessments Arrive to lecture and lab on time and be active participants Create relevant assessments and class activities
Respect	 Speak openly with others while honoring diverse viewpoints and perspectives Allow others to voice their opinions and perspectives 	 Respect your perspective even while we challenge you to think more deeply and critically Help facilitate the respectful exchange of ideas in class
Fairness	 Contribute fully and equally when working in teams Not seek unfair advantage over others 	 Create fair assessments and grade in a fair and timely manner Treat students and teams equally
Trustworthy	Not engage in personal	Be available when we say we

	affairs while on class time Be open and transparent about what you are doing in class Not distribute course materials to others in an unauthorized fashion	 will be Follow through on our promises Not modify the expectations or standards without communicating with everyone
Courage	 Say or do something when you see actions that undermine any of the above values 	Say or do something when we see actions that undermine any of the above values

This class statement and table of values is adapted from Dr. Tricia Bertram Gallant.

All course materials are the property of the instructor, the course, and the University of California, San Diego and **may not** be posted online, submitted to private or public repositories, or distributed to unauthorized people outside of the course. Any suspected instances of a breach of academic integrity will be reported to the Academic Integrity Office for review. A breach of academic integrity may result in a zero on the assignment/test/participation item in question and/or a failed grade in the course.

If you observe anyone not acting in accordance with the above values we are trying to foster, please bring your concerns to my or the instructional team's attention, and we will do our best to determine appropriate actions to uphold and protect these values.

Submitting reports to Turnitin.com: Students agree that by taking this course all required papers will be subject to submission for textual similarity review to Turnitin.com for the detection of plagiarism. All submitted papers will be included as source documents in the Turnitin.com reference database solely for the purpose of detecting plagiarism of such papers.

Letters of Recommendation:

If you think you may want me to write you a letter of recommendation (or any other instructor), please consider what a good letter would contain and how your actions in the course demonstrate the qualities you will want highlighted in a good letter. When students ask me for a letter of recommendation, I ask them to write to me about how they demonstrated critical thinking, leadership, collaboration, and professionalism. I will be specifically looking for examples of these qualities that I could have noticed during lecture, lab and office hours. Be sure to actively participate in the discussions, talk to me during the lab and my office hours: ask questions, offer your own ideas and interpretations of your results, bring interesting facts/papers that are connected to the material we are studying. If you don't actively show the qualities that are needed to write a good letter, it will be hard for me to write a letter that is meaningful and useful.

Tentative Course Schedule: (subject to change)

	Dates	Lab activities: "Wet" = in-person, "Zoom" = online, real-time lab	Relevant Lab Manual Sections
Week 1	Tues/Wed	Cimile, real anne las	How to Use the Manual; Working in the Lab; Keeping a Good Lab Notebook; Safety Rules; Instructions for disposal of laboratory waste; BACKGROUND: Pipette Operation; Liquid Measurement Units, Basic Dilutions, Serial Dilutions; PROTOCOLS: APPENDIX G,
NOOK 1	Jan 4/5	Wet: Intros, safety, dilutions	C, D
	Thur/Fri Jan 6/7	Zoom: molecular biology boot camp (review)	APPENDIX H
Week 2	Tues/Wed Jan 11/12 Thur/Fri	Wet: Extracting pML104 plasmid, running agarose gel to check extractions Zoom: Bioinformatics - Exploring the ADE2 gene to	BACKGROUND: Cultures used in the lab; Plasmids used in the lab; Alkaline Lysis Plasmid Purification; Spectrophotometric Analysis of DNA & RNA; Agarose Gel Electrophoresis; PROTOCOLS: 3 (for reference: cultures will be set up ahead of time); Protocols 4, 5, 6 for doing in the lab BACKGROUND: CRISPR-Cas9 Project Overview; PROTOCOLS: 1,
	Jan 13/14	identify important features and where to mutate	APPENDIX B
Week 3	Tues/Wed Jan 18/19	Wet: Restriction enzyme digestion of pML104 plasmid, check digestions with agarose gel electrophoresis, clean digested plasmid for future use in ligation	BACKGROUND: Restriction Enzyme Cloning; PROTOCOLS: 7, 6, 8, 5 BACKGROUND: CRISPR-Cas9 Project Overview - editing the ADE2
	Thur/Fri Jan 20/21	Zoom: CRISPR experimental design; designing gRNA & HDR template sequences	gene + Homology Directed Repair of ADE2; <u>PROTOCOLS</u> : APPENDIX J; PROTOCOL 2 (2a and 2b).
	3411 Z0/Z1	Week 4 *Double We	
	Tues/Wed	Wet: Plan & set-up ligations; bacterial	BACKGROUND: Restriction enzyme cloning; Ligation; Annealed
	Jan 25/26	transformation with ligations	Oligo + Restriction Enzyme Cloning; PROTOCOLS: 9, 10
Week 4	Thur/Fri Jan 27/28	Wet: Analyze ligation-transformation results; CRISPR experimental design	PROTOCOL: 10 (Analyzing E. coli transformations)
		Week 5 *Double We	et lab week*
Week 5	Tues/Wed Feb 1/2	Wet: Set-up HDR PCR, check via agarose gels, and clean; set up overnight cultures of pML104- gRNA	BACKGROUND: Polymerase Chain Reaction (PCR); Making Copies of HDR templates; PROTOCOLS: 14, 6, 7, 15 - Part 1
	Thur/Fri Feb 3/4	Wet: extract pML104-gRNA plasmids and check using agarose gels; re-streak yeast to grow a fresh plate for transformations in week 6	
		Week 6 *Double We	et lab week*
	Tues/Wed Feb 8/9	Wet: Yeast transformations	PROTOCOLS: 15 - Part 2 through step 7
Week 6	Thur/Fri Feb 10/11	Wet: Plate yeast tranformations; Journal club (DiCarlo et al. 2012 paper)	PROTOCOL: 15 - Part 2, step 8-11; APPENDIX I
		Week 7 *Flip week - Wet	lab on Thurs/Fri*
	Tues/Wed		
Week 7	Feb 15/16 Thur/Fri	Zoom: Analyze yeast transformations Wet: Count yeast colonies; Extract genomic DNA	BACKGROUND: Polymerase Chain Reaction (if refresher needed);
Week 7	Thur/Fri Feb 17/18 Tues/Wed	Wet: Count yeast colonies; Extract genomic DNA from yeast cultures, set-up ADE2 PCR Wet: Run PCRs on agarose gels, clean and send	PROTOCOLS: 16, 17 (step 1 only)
Week 7	Thur/Fri Feb 17/18	Wet: Count yeast colonies; Extract genomic DNA from yeast cultures, set-up ADE2 PCR	
	Thur/Fri Feb 17/18 Tues/Wed Feb 22/23 Thur/Fri	Wet: Count yeast colonies; Extract genomic DNA from yeast cultures, set-up ADE2 PCR Wet: Run PCRs on agarose gels, clean and send for sequencing	PROTOCOLS: 16, 17 (step 1 only) PROTOCOLS: 17 steps 2-4 BACKGROUND: Sanger DNA sequencing; PROTOCOLS: 18, Part 1 and 2
	Thur/Fri Feb 17/18 Tues/Wed Feb 22/23 Thur/Fri	Wet: Count yeast colonies; Extract genomic DNA from yeast cultures, set-up ADE2 PCR Wet: Run PCRs on agarose gels, clean and send for sequencing Zoom: Sequence analysis	PROTOCOLS: 16, 17 (step 1 only) PROTOCOLS: 17 steps 2-4 BACKGROUND: Sanger DNA sequencing; PROTOCOLS: 18, Part 1 and 2
Week 8	Thur/Fri Feb 17/18 Tues/Wed Feb 22/23 Thur/Fri Feb 24/25 Tues/Wed March 1/2	Wet: Count yeast colonies; Extract genomic DNA from yeast cultures, set-up ADE2 PCR Wet: Run PCRs on agarose gels, clean and send for sequencing Zoom: Sequence analysis Week 9 *Double Wet Review of overall data set; DNA Extraction & PCR for TAS2R38/PTC project	PROTOCOLS: 16, 17 (step 1 only) PROTOCOLS: 17 steps 2-4 BACKGROUND: Sanger DNA sequencing; PROTOCOLS: 18, Part 1 and 2 It lab week*
Week 8	Thur/Fri Feb 17/18 Tues/Wed Feb 22/23 Thur/Fri Feb 24/25 Tues/Wed March 1/2 Thur/Fri March 3/4	Wet: Count yeast colonies; Extract genomic DNA from yeast cultures, set-up ADE2 PCR Wet: Run PCRs on agarose gels, clean and send for sequencing Zoom: Sequence analysis Week 9 *Double Wet Review of overall data set; DNA Extraction & PCR for TAS2R38/PTC project Wet: Digestion of TAS2R38 PCRs, PTC taste-test,	PROTOCOLS: 16, 17 (step 1 only) PROTOCOLS: 17 steps 2-4 BACKGROUND: Sanger DNA sequencing; PROTOCOLS: 18, Part 1 and 2 et lab week* APPENDIX L, including Protocols 23 & 24 APPENDIX L, including Protocols 25-28
Week 8	Thur/Fri Feb 17/18 Tues/Wed Feb 22/23 Thur/Fri Feb 24/25 Tues/Wed March 1/2 Thur/Fri	Wet: Count yeast colonies; Extract genomic DNA from yeast cultures, set-up ADE2 PCR Wet: Run PCRs on agarose gels, clean and send for sequencing Zoom: Sequence analysis Week 9 *Double Wet: Review of overall data set; DNA Extraction & PCR for TAS2R38/PTC project Wet: Digestion of TAS2R38 PCRs, PTC taste-test, start analyzing data; Review of CRISPR write-up	PROTOCOLS: 16, 17 (step 1 only) PROTOCOLS: 17 steps 2-4 BACKGROUND: Sanger DNA sequencing; PROTOCOLS: 18, Part 1 and 2 et lab week* APPENDIX L, including Protocols 23 & 24 APPENDIX L, including Protocols 25-28

Potentially Useful Resources (listed alphabetically):

Black Resource Center: The Black Resource Center is a Campus Community Center that serves everyone at UC San Diego while emphasizing the Black experience. We promote scholarship, foster leadership, and cultivate community for students through the committed, collaborative effort and support of faculty, staff, and the broader UC San Diego community.

http://brc.ucsd.edu/ (website)

Counseling And Psychological Services (CAPS): CAPS provides FREE, confidential, psychological counseling and crisis services for registered UCSD students. CAPS also provides a variety of groups, workshops, and drop-in forums. http://caps.ucsd.edu/ (website)

Cross-Cultural Center: The Cross-Cultural Center strives for meaningful dialogues and context across all cultures, particularly those of underrepresented or underprivileged backgrounds. We offer supportive and educational services through art, social and educational programs, workshops, and outreach. We welcome creative venues for enhancing social consciousness and equity. http://ccc.ucsd.edu/ (website)

Hub Basic Needs Center & Triton Food Pantry: The Hub Basic Needs Center addresses the gaps and concerns students have with accessing nutritious food, stable housing, and financial wellness resources. We are committed to transforming dialogues surrounding the basic needs of students so they can focus on their academic success contributing to holistic well-being.

https://basicneeds.ucsd.edu/ (website)

Inter-Tribal Resource Center: We are focused on supporting Native American students and promoting educational access in our tribal communities. https://itrc.ucsd.edu/ (website)

LGBT Resource Center: The Lesbian Gay Bisexual Transgender Resource Center at UC San Diego provides a visible presence on campus and enhances a sense of connection and community among LGBT faculty, staff, students, alumni and the UC San Diego Community.

http://lgbt.ucsd.edu/ (website)

Office for Students with Disabilities (OSD): The Office for Students with Disabilities (OSD) at UC San Diego works with undergraduate, graduate, and professional school students with documented disabilities, reviewing documentation and determining reasonable accommodations.

https://disabilities.ucsd.edu/about/index.html (website)

Raza Resource Centro: The Raza Resource Centro team is committed to our student's success and we work collectively to meet the needs of our students. The Centro is a lively space where students study, meet, write, get tutoring, and most importantly are in community. It is a space where Latina/o Chicana/o organizations hold meetings, events and where culture, arte, and academics interconnect. http://raza.ucsd.edu/ (website)

Student Veterans Resource Center: The Student Veterans Resource Center (SVRC) is committed to ensuring that military affiliated students successfully make the transition from the military environment to campus life, and are assisted in their progress toward completing their academic degree. The Center also provides opportunities for peer-to-peer support, mentoring and social networking. https://students.ucsd.edu/sponsor/veterans/ (website)

Women's Center: The Women's Center serves as a resource for the entire campus community while placing the experiences of diverse women at the center through the resources we provide, the programming and learning opportunities we facilitate, and the dynamic community space that we create. https://women.ucsd.edu/ (website)

There are many other resources available to you on campus, and if you wish to know more about where you can go for support – please let me know and we can find it together. If you would like me to include resources other than those I have listed above, let me know as well! I want to know what is important for everyone!