

# BIBC 103: Biochemical Techniques

Winter Quarter, 2022

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## Course Shifted to Remote Format Through Week 4

The first two weeks of the class will be conducted remotely. Lectures, lab sessions, and office hours will be conducted by Zoom. Log into Canvas and join through the link on the Zoom LTI page or in the Calendar. The lectures will be recorded, but the lab sessions will not be recorded. You are required to attend lab on Zoom with your video on (please contact me if you are not set up for this). You will work in groups of four to complete the lab work, and during the lab sessions you will go back and forth between your lab class of 24, and breakout rooms where you will work in your lab groups. The lab sessions will not be recorded. Some of the lab work is for credit for what is completed during the lab sessions. In week 5, the class will shift to an in-person format for lectures, lab, and office hours. Lectures will be video-recorded and made available by podcast. There will not be a remote option for the labs after the first two weeks, and attendance in person will be mandatory.

**Office Hour:** Tata Hall 3102 conference room, Mondays 2 – 3 pm

**Lecture:** Mon/Weds/Fri 9 – 9:50 am in Center Hall 212. You are strongly encouraged to attend lecture. While it is not mandatory, in lecture we will discuss the background to the labs and strategies for approaching the lab work and assignments. The lectures will be recorded for video podcast, but attending live gives you the opportunity to ask questions.

**Labs:** York Hall 3306 and 3406; Weds/Fri 10 am – 2 pm or 2:30 – 6:30 pm.

## **Course Objectives:**

This course will introduce some of the experimental methods used in biochemistry and molecular biology, with an emphasis on those techniques used to study proteins. You will gain conceptual understanding and hands-on experience in various protein purification techniques and methods for analyzing the different properties of proteins. The laboratory work will consist of three big multi-week projects and some shorter side projects. All the lab work will emphasize mastery of the skills that are essential to work independently in a biochemistry lab, including hands-on wet-lab and quantitative reasoning skills.

More importantly, this course is designed to give an appreciation of what science is and how it works. Science is not just a bunch of random facts...it is a process! It is easier to understand biology, or any field, when you understand how we know what we know about it. Understanding how information in biology is brought to light is just as important as the information itself. Through the laboratory projects we will develop the skills necessary to interpret data from experiments in order to answer questions about biological systems, and to

design experiments to ask new questions. In keeping with this, the importance of good experimental design, including the use of appropriate controls, will be highlighted in all experiments.

**Materials Required:**

- 1) Biochemical Techniques Lab Manual, 2020 – 2021 Edition (available from the Bookstore)
- 2) Bound laboratory notebook (see Canvas for information on the type of notebook required)
- 3) Safety glasses
- 4) Lab coat

**Course Structure**

Activity	Point Value
LDH Purification: Lab notebook	40
LDH Purification: Purification table analysis	115
FGF Signaling: Questions from lab manual data (Jan 14)	10
FGF Signaling: Lab report	230
Bioinformatics: Lab manual questions	60
Bioinformatics: PyMOL protein image	50
Fluorescent Proteins: Lab notebook	35
Fluorescent Proteins: Identify unknowns	20
Lab 1 part E calculations (Jan 5)	10
Quizzes: Three at 30 points each	90
Exam 1	130
Exam 2	210
<b>Total</b>	<b>1000</b>

**Point Cutoffs for Grade Assignments:** (Cutoffs may be lowered at the instructor’s discretion.)

910-1000	A	790-799	C+
900-909	A-	705-789	C
890-899	B+	695-704	C-
810-889	B	600-694	D
800-809	B-	0-599	F

**Course Web Site:**

Except for the lab manual, all course materials will be accessed through the course webpage on [Canvas](#). Much of the data you generate in your experiments will be accessed through Canvas, in addition to lab report guidelines and practice problem sets for quizzes and exams. Be sure to check Canvas frequently for announcements and updates on assignments.

### **Lab Quizzes and Exams:**

All quizzes and exams will be taken in person during the lab sessions. The purpose of the lab quizzes is to be sure you are mastering the basic concepts behind your experiments as we go through the class. This includes understanding the purpose of the lab projects and how each experiment fits into this, the basic concepts underlying the procedures, and simple mathematical and analytical skills based on what you have actually done in lab. The quiz dates are given in the lab schedule. Quizzes will be given at the beginning of lab, will take 15 – 30 minutes, and will consist of 5 to 7 questions. The topics that will be covered on each quiz will be posted in an announcement on Canvas on the Monday prior to the quiz.

The two exams are cumulative and will be problem solving-based. They may include some basic questions on the concepts we have covered, but will emphasize taking the information you have learned and extrapolating to solve problems you have not seen before. Practice questions will be provided on Canvas to help you prepare for the exams.

### **Lab Attendance Policies:**

In-person attendance at each lab session is mandatory. If you test positive for COVID-19 or feel ill, however, stay home and contact the instructor by email. If you test-positive for COVID-19 and must isolate, we will work with you to keep you in the class. If you know that you need to miss a lab session, discuss this with the instructor (not the IA, they are not authorized to give you permission) to see if it will be possible to make up the lab session or excuse you from the lab with no consequences. Please bring this to the instructor's attention as soon as you know that it will be an issue. An unexcused absence will result in 10 points being deducted. **Only the instructor can excuse an absence. Two unexcused absences will result in the student failing the course.**

### **Turning Written Assignments:**

Lab report and data write-ups will be submitted electronically on Canvas. Lab reports are due before the end of the day (11:59 pm) on the due date. Ten points will be deducted for each day following the due date that the lab report is late. Students agree that by taking this course all required papers will be subject to review for textual similarity by Turnitin.com for the detection of plagiarism. All submitted papers will be included as source documents in the Turnitin reference database solely for the purpose of detecting plagiarism of such papers. Use of the Turnitin service is subject to the terms of use agreement posted on the Turnitin site.

**Lab Report Grading and Regrade Policy:** Your lab report will be graded by your IA, based on the lab report guidelines. I work closely with all the IAs to ensure that the grading is accurate and equivalent between sections. If you disagree with the grading of your lab report, discuss this with your IA to get clarification on why points were deducted. If you still disagree with the grading you may submit the report to me for a re-grade. This must be done within one week of receiving the graded report. I will re-grade the entire report and give you a new score, and this is the score that will be recorded.

## Lab Schedule

	Dates	Experiment/Activity	Lab Manual Chapter
<b>Wk 1 Remote</b>	Jan. 5	Arrange groups; Pipetting video and convert absorbance to percent concentration to pipetting accuracy	Lab 1 part E
	Jan. 7	Bioinformatics 1	Lab 19 part A
<b>Wk 2 Remote</b>	Jan. 12	Bioinformatics 2	Lab 18 part A Lab 19 parts C and D
	Jan. 14	Bioinformatics 3 (select proteins for PyMOL protein image project)	Lab 19 part D
<b>Wk 3 Remote</b>	Jan. 19	Fibroblast Growth Factor (FGF) Signaling: Examine dataset, come up with questions <b>Quiz 1—Asynchronous Canvas quiz</b>	Lab 9B
	Jan. 21	FGF 2: Develop hypotheses to explain dataset and design experiments to test	Lab 9B
<b>Wk 4 Remote</b>	Jan. 24	PyMOL protein image assignment due Mon by 11:59 pm	
	Jan. 26	FGF 3: Examine Western blot and ELISA data	Labs 10 - 12
	Jan. 28	FGF 4: Design figures for lab report; Electrophoresis & SDS-PAGE—calculations for SDS-PAGE	Labs 10 – 12 Lab 2
<b>Wk 5</b>	Feb.2	Safety orientation; Electrophoresis & SDS-PAGE—run SDS-PAGE	Lab 2
	Feb. 4	LDH 1: Initial purification of LDH from crude homogenate: centrifugation, ammonium sulfate precipitations	Lab 3
<b>Wk 6</b>	Feb. 9	Exam 1 in lab; prepare size exclusion columns	Lab 5
	Feb. 11	LDH 2: Affinity chromatography	Lab 4
		FGF signaling lab report due Friday by 11:59 pm	
<b>Wk 7</b>	Feb. 16	LDH 3: Size exclusion chromatography	Lab 5
	Feb. 18	LDH 4: LDH activity assays; Bradford protein assays	Lab 6
<b>Wk 8</b>	Feb. 23	LDH 5: SDS-PAGE of LDH purification fractions <b>Quiz 2</b>	Lab 7
	Feb. 25	LDH 6: Agarose gel electrophoresis of LDH isozymes	Lab 8
<b>Wk 9</b>	Feb. 28	LDH purification table analysis due Mon by 11:59 pm	
	Mar. 2	Fluorescent proteins (FP): Make competent cells and transform with plasmid <b>Quiz 3</b>	Lab 15
	Mar. 4	Purification and analysis of fluorescent proteins	Lab 16
<b>Wk 10</b>	Mar. 9	SDS-PAGE of fluorescent proteins	Lab 17
	Mar. 11	Exam 2 in Lab	