BIMM 101 Recombinant DNA Techniques Spring 2012

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Lectures: Tue, Thu 8 – 9:15 AM CSB 001

Labs: D01 Tue / Thu 9:30-1:30, or Wed / Fri 9 – 1 York 4318, 4332

Course Website: http:// ted.ucsd.edu

- Lectures will cover the theory behind the experiments performed in lab. The quizzes and exam will have questions on the material that will be covered during lectures.

Learning objectives:

- Learn the theory behind molecular techniques, and the applications of the methodologies in biological research
- · Become proficient at basic molecular biology techniques
- Learn the importance of proper controls in designing experiments and interpreting results
- Improve lab math skills and ability to graph data correctly
- Learn to make logical conclusions from experimental data
- Become familiar with bioinformatics databases and applications
- Learn to find, read, and evaluate primary literature
- Become aware of the implications of the technology for society

Required texts:

BIMM 101 Lab Manual from University Readers From Genes to Genomes by Dale (1st or 2^{cd} edition) on reserve at BML and electronic version available from UCSD computer http://onlinelibrary.wiley.com/book/10.1002/0470856912 Readings on TED (ted.ucsd.edu)

Required Materials – needed by second day of class:

Labcoat (the bookstore has cheap ones)
UV blocking safety glasses (also at bookstore)
Lab notebook with carbon copies (bookstore or Grove general store)
Fine point Sharpie for labeling – get a dark color
A calculator

Remember that lab attendance is required – if you miss two labs, you will be asked to drop the course. If you are ill, you must leave a message with your instructor, not your TA, and make up the lab in a way that we will determine. You must be on time for lab; the TAs go over the experiments at the beginning of lab, and also quizzes are administered then.

Course Requirements

- **1. Lab Notebook**: It is mandatory that you keep a complete lab notebook. The notebook must contain everything that you did in the lab, including:
- Any changes in the protocol
- All data/results
- All calculations done during experiments
- Observations
- **2. Take home assignments:** there will be two take-home assignments
 - dilutions 10 pts
 - DNA quantification 20 pts
- **3. Quizzes**: There will be 4 scheduled quizzes during class periods staring week 3, each worth 10 points. They will be given at the beginning of class, and collected 15 minutes later. If you arrive after the quiz has been handed out, you will not be able to make it up.
- **4. Lab Reports**: 4 lab reports throughout the quarter are worth 40 points each. While you will be collecting and sharing data with a lab partner, and you are welcome to discuss your results with your classmates, you must hand in your own lab report, written in your own words. You will be penalized for copying another lab report or for handing in the same (or very similar, such as just a few words changed here and there) lab reports as your partner.

All lab reports should include:

- 1. Purpose of the experiment: this section should be BRIEF no more than a few sentences: simply state why you are doing the experiment.
- 2. Results should include the following:
- data or data analysis
- figures, gels (or representations thereof)
- any sample calculations
- a brief statement about what each result means
- 3. Discussion:
- Note any unusual observations
- Discuss success or failure of the experiment if there was a problem, discuss probable source.

Lab reports should have no more than 5 pages of text – figures can go on

separate pages. More specific description of the lab report can be found in the course TED page. Although the lab report will be submitted electronically, all carbons from the labs associated with a lab report must be handed in class the day the lab is due.

Lab report Late policy.

Lab reports are due at the beginning of lecture on the assigned due date (must be electronically submitted before 11 AM of the due date).

Penalty for turning lab reports late:

- 5 points if handed in later on the same date;
- After the first late day, you lose 2 points/day, so
- -7 points if handed in anytime the next day
- -9 points if handed in the 3rd day etc.

Lab report due dates:

Labs to be included in the Lab Report	Due date	
Labs 3-8A,	Tue, May 01	Lab Report #1
Labs 9-15A, excluding 14c	Tue May 22	Lab Report #2
Labs 16- 18A	Thu, June 1	Lab report #3
Labs 8B, 14C, 15B, 18B, 19	Thu, June 7	Lab Report #4

- **5. Lab attendance**: Attendance is taken within the first 15 minutes of every lecture session. If you are ill, please notify me (gbozinovic@ucsd.edu).
- **6. Lab performance:** There are no points for lab performance per se. However, your effort, attitude, and the success of your experiments will be considered when assigning the final grade, especially if you are on the borderline between two grades. If you miss one lab with no excuse, you will lose 5% from your final grade. If you miss two labs, you will receive an F for the course.
- **7. Exam**: There will be one cumulative exam worth 50 points. Depending on the performance of the class, the exam scores might be adjusted.
- The exam adjustment policy:
- 1) If any student receives a 100% on the exam it will NOT be curved;
- 2) If any student receives a score between 95 and 100%, that will be the new maximum score (for example if the highest grade is 96, everyone's score will increase by 4 points);
- 3) If the highest score is less that 95% then that student's score will be the new 95% (for example if the highest grade is 89, everyone's score will increase by 6 points).

Extra Credit: Students are encouraged to present the topic of their choice relative to class material in a short, 5 min presentation. Students may present individually, or in a groups of two. There is no specific format required. You can earn up to 16 pts (5%) extra toward your final grade.

Grading:

Attendance (2 points per lab)	36 pts
Dilution assignment	10 pts
Lab 2 (DNA quantification) assignment	20 pts
4 Quizzes (10 points each)	40 pts
4 Lab reports (40 points each)	160 pts
Final Exam	50 pts

Total possible points: 316

Please make sure you regularly check your scores in WebCT to make sure no errors have occurred.

Letter grades will be assigned as follows: Grade Overall class percentage A, A->92, 90 B+, B, B- 88, 82, 80 C+, C, C- 78, 72, 70 D+, D, D- 68, 62, 60 F Below 60

Note: Just coming to lab does not ensure that you will get a passing grade in the class. You must hand in all assignments and get passing scores on those assignments (an average of 70) to get a C- in the class.

Policy on cheating: Anyone caught cheating (which includes, but it is not limited to plagiarizing lab reports, cheating on a test or quiz, or changing an answer for a regrade) will be reported to the Academic Integrity Office.

Letters of recommendation: If you think you need a letter of recommendation at some point in the future, please save our lab reports and tests, as I will ask for some of them to review. Also, I will write letters only for those who receive an A in the course. Finally, even if you have an A, if you have never spoken to me or come to my office hours, I may not agree to write a letter for you.

BIMM 101 Spring 2012 Student contract:

1. I understand that if I am late for lab or allowed to take the quiz and will receive	• •
Name	Date
work in your own words. While discussion encouraged, each student on their own mustigures, graphs, and tables. The submission contain shared work is forbidden, and will reboth reports. The exception to this is when supplied to each member of the group (spendotographs). In this case the labeling of the independently. If you have questions about your work with others and unauthorized color T.A. for clarification.	st complete all text, references, in of reports by lab partners that esult in points being deducted from a figure is the raw data that is cifically absorption spectra and gelenat figure must be done to the difference between discussing laboration, please ask your instructor own work in your own words, you may atory reports that were written by in, which is a direct attempt by the where own, and is no different than terial from other sources without som, even if the source is cited as a rously sought out and penalized. This of plagiarism checker against all this of plagiarism will automatically be ator. Following UCSD's Policy on edu/manual/appendices/app2.htm), in or other academic misconduct will the Council of Deans) and academic more, all submitted reports are whits by the plagiarism checker will ovided the plagiarized material. Giving low them to copy material from that I be pursued and penalized as a plagiarism. The port and it is detected by cademic Integrity Office on a lab report to a student who takes or she plagiarizes my lab report, I
Name	 Date

Tentative Schedule

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Lab 1	A. PipettingB. DilutionsC. Bacteriological TechniquesD. Calibration of a pipettemen
Lab 2	A. Agarose Gel Electrophoresis
Lab 3	 A. Introduction to bioluminescence B. Course Objectives C. P1: Isolation of chromosomal DNA from <i>Vibrio fischeri</i>
Lab 4	A. P2: Isolation of chromosomal DNA from Vibrio fischeri
Lab 5	 A. Spectrophotometric Analysis of Vibrio DNA B. Sal I restriction digestion of Vibrio fischeri and pGEM DNA
Lab 6	 A. Agarose gel electrophoresis of Sall restriction digest and estimate of DNA quantities from gel B. Ligation of <i>Vibrio fischeri</i> DNA with pGEM DNA
Lab 7	 A. Ligation assessment by gel electrophoresis B. Transformation of competent E. coli DH5α cells
Lab 8	A. Count colonies and re-streak glowing colonies from original transformationB. Isolation of DNA from bees
Lab 9	 A. Amplification of <i>lux</i>AB gene by PCR B. Start overnights for experiment to size and seq. <i>V. fischeri Sal</i> I inserts C. Bioinformatics computer Lab 1
Lab 10	 A. PCR success assessment by gel electrophoresis B. Alkaline lysis miniprep: purification of plasmid DNA C. Restriction Digest with <i>Sal</i> I D. Clean up PRR Products
Lab 11	 A. Run gel on Sal I restriction digest products B. Restriction digest of lux plasmids, PCR products; Cloning vector pUC18 for subcloning luxAB

Lab 12	 A. Agarose Gel electrophoresis of EcoRV/Sac I digest from Lab 9 B. Cut band out of gel and purify DNA from agarose for Cloning C. Case it! Restriction digest and Southern blot simulation
Lab 13	 A. Quantification of DNA from gel B. Subcloning luxAB into a plasmid vector-ligation C. Bioinformatics Lab 2
Lab 14	 A. Making competent DH5α cells B. Transformation of ligation products into DH5α cells C. Run gel of plant PCR products and clean up
Lab 15	 A. Screening for colonies containing <i>lux</i>AB by adding exogenous aldehyde B. Analyze plant sequences
Lab 16	A. Set up C. elegans plates for RNAi experiment
Lab 17	A. Observe worm phenotypes and isolate RNAB. Quantitate RNA and set up quantitative RTPCR
Lab 18	A. Analyze RTPCR resultsB. Isolate cheek cell DNA and set up PCR for PTC
Lab 19	A. PTC taste test and PCR product digestB. Run Gel to PTC PCR product and analyze resultsC. Review

Exam and Quiz Schedule:

Quiz 1 Tue, Apr 17 Quiz 2 Thu, May 3 Quiz 3 Thu, May 17 Quiz 4 Tue, May 29

FINAL EXAM: Thursday, June 7th