### **WELCOME TO BIMM 101, FALL 2018, UC SAN DIEGO**

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OFFICE: 4121 Bonner Hall (Lab)
OFFICE HOURS: TBD, **Bonner Hall 4121**.

I also strongly encourage you to find me during lab time.

INSTRUCTIONAL ASSISTANTS: B01 Selena Chen syc011@ucsd.edu

B02 Jay Chitale <u>ichitale@ucsd.edu</u>

LECTURE: 10:00 – 10:50 AM MWF CSB 005

LABS: 11:00 AM – 2:50 PM WF B01: YORK 4318

B02: YORK 4332

COURSE WEBSITE: <a href="http://tritoned.ucsd.edu">http://tritoned.ucsd.edu</a>

REQUIRED MATERIALS -- bring to lab each day, required by second day of lab:

- 1. Labcoat must go to knees (available at bookstore)
- 2. UV blocking safety glasses (also at bookstore)
- 3. BIMM 101 Lab manual (available at bookstore)
- 4. Carbonless copy laboratory notebook (available at bookstore)
- 5. Fine point Sharpie for labeling get a dark color
- 6. Calculator you cannot use a cell phone for quizzes!
- 7. iClicker (available at bookstore, version 2 preferred-register on TritonEd)
- 8. Long pants and closed-toed shoes are always required in lab (entire legs and feet covered)

RECOMMENDED TEXT – From Genes to Genomes by Dale. On reserve at BML.

Electronic version available at (download from UCSD computer):

http://onlinelibrary.wiley.com/book/10.1002/0470856912

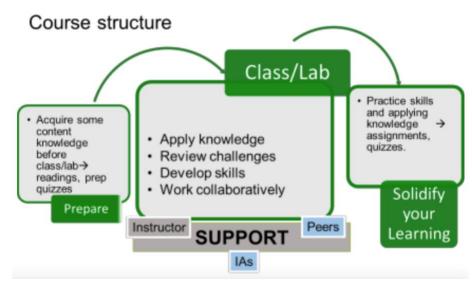
PURPOSE OF THIS COURSE – to develop an understanding of research in molecular biology through inquiry-based laboratory experiments. Students will work in groups to collect, analyze, and present research data while learning molecular and biological concepts and laboratory skills.

### **LEARNING GOALS:**

- Apply knowledge of molecular biology concepts and molecular techniques to plan experiments, explain and troubleshoot results
- Demonstrate proficiency at the basic molecular biology techniques used in the lab
- Explain the importance of proper controls in designing experiments and interpreting results
- Perform basic lab math skills, statistical analysis, and graphing
- Draw conclusions based on evidence and reasoning
- Use basic bioinformatics databases and applications
- Find, read, and evaluate primary literature
- Collaborate with one another to learn foundation biological concepts and laboratory skills

## LEARNING IN THIS COURSE

This course is designed to be a collaborative environment for everyone to learn together and construct a shared understanding of the material. Active participation both in class and lab is expected. Being able



to communicate understanding, and confusion, is critical to success in any discipline, and is very useful for learning<sup>1</sup>. To encourage communication and collaboration, we will frequently use class time to work on problems in groups.

We like to use class time to work on applying knowledge, troubleshooting difficult topics, and practice solving problems. Hence, it is expected that you will prepare before coming to class, reviewing basic background information about the lab and/or relevant content. This will be encouraged through targeted readings and in-class quizzes. The more prepared you are for class and lab, the more fruitful our discussions can be.

Instead of memorization, we will focus on developing an understanding of fundamental concepts and as they apply to the experiments. Therefore, tests will include questions that are based on solving problems in new contexts or data interpretation and not necessarily on memorizing facts.

1 Smith et al., 2009. http://www.sciencemag.org/content/323/5910/122.short

#### ACCESSIBILITY AND INCLUSION

http://disabilities.ucsd.edu | osd@ucsd.edu | 858-534-4382

Any student with a disability is welcome to contact us early in the quarter to work out reasonable accommodations to support their success in this course. Students requesting accommodations for this course due to a disability must provide a current Authorization for Accommodation (AFA) letter issued by the Office for Students with Disabilities (OSD). Students are required to present their AFA letters to faculty and to the OSD Liaison in the Division of Biological Sciences in advance so that accommodations may be arranged.

Whenever possible, we will use universal designs that are inclusive. If you have feedback on how to make the class more accessible and inclusive, please let us know!

GRADING -- BIMM101 has four grading components: participation & professionalism (20%), quizzes (35%), laboratory reports (35%). Because different people may excel in different aspects, the laboratory reports or quizzes, whichever is higher for each individual, will be scaled to 45% instead of 35%, bringing the total to 100%. The following grading scheme will be used.

A+	97-100%	B+	87-90%	C+	77-80%	D+	67-70%	F	0-60%
А	93-97%	В	83-87%	С	73-77%	D	63-67%		
A-	90-93%	B-	80-83%	C-	70-73%	D-	60-63%		

The course is not graded on a curve (i.e. 20% of students getting A, B, C, and such). Thus, the ability to do well in this course is not dependent on others doing poorly.

There are no opportunities for extra credit beyond what is assigned as part of the course by the instructor.

PARTICIPATION AND PROFESSIONALISM (20%) – Success in this course depends upon the willingness of student to engage in both individual and team-oriented activities that foster the mindset of scientific research environments. This portion of the grade consists of the following:

pre-lecture activities (6%) – will include but not be limited to reading assignments, quizzes, or problem sets. Material for these activities will be posted on TritonEd.

lecture activities (5%) – will occur on a regular basis and will include i>clicker questions.

- pre-laboratory activities (2%) prior to coming to lab, students are to create a flow chart outlining the procedures that will be performed in lab that day, written on the carbon paper from the lab manual. The flow charts will be collected during the first 10 minutes of the lab.
- laboratory notebooks (5%) dedicated, professional record keeping of work done in the laboratory is critical to success in this course and your future career. On the lab report due dates students will submit the carbon copies of the relevant notebook sections. In addition, notebooks will be subject to spot-checking during the course of the quarter.
- professionalism (2%) The laboratory environment is a collegial, collaborative, professional community and student behavior toward their peers and instructors is expected to reflect as much. This portion of the grade reflects upon both the actions of the student as an individual and the class as a whole. As such 1% accounts for the initial assumption of each student as dedicated, collaborative professionals, and chronic deviations of individual students will result in points being deducted. The remaining 1% reflects upon the class community and refers to surveys and evaluations. If 90% of the class responds to such material designed to provide feedback for the improvement of the course and the instructors and assistants, all students will earn these points.

QUIZZES (35%) – There will be a total of 5 quizzes throughout the quarter, with the dates noted on the course schedule. Each quiz will be cumulative but will emphasize the most recent material from both lecture and laboratory sessions. The first 4 quizzes are each worth 5%, with the final quiz worth 15% of the total grade.

LABORATORY REPORTS (35%) – Students will submit a total of 5 mini reports describing the results of their laboratory work. Specific instructions will be posted on TritonEd, but they will generally follow the format of peer-reviewed scientific research publications (you can use Saltman Quarterly as a guide).

- Gel Electrophoresis Report 5%
- PCR Variations Report 8%
- Ligation Efficiency Report 10%
- Promoter Mutants Report 12%

These reports are to be submitted electronically over TritonEd through TurnItIn before coming to lab. In addition, at the beginning of lab you will turn in a hard copy with the carbon sheets from the relevant notebook pages attached.

EXPERIMENTAL SUCCESS – student grades do not depend upon whether the experiment "worked," but instead upon the critical analysis of the results (or lack thereof). Recognizing where problems in the experiment arose, and presenting evidence to support your conclusions is a critical part of being an successful scientific researcher. Note, however, that chronic carelessness is still noted in the Participation and Professionalism portion of the grade.

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# ACADEMIC INTEGRITY -- (<u>https://students.ucsd.edu/academics/academic-integrity/index.html</u>)

Integrity of scholarship is essential for an academic community. The University expects that both faculty and students will honor this principle and in so doing protect the validity of University intellectual work. For students, this means that all academic work will be done by the individual(s) to whom it is assigned, without unauthorized aid of any kind. Anyone caught cheating (includes plagiarizing lab reports, cheating on a test, or changing an answer for a regrade) will be reported to the Academic Integrity Office.

All course materials are the property of the instructor, the course, and the University of California, San Diego and may not be posted online, submitted to private or public repositories, or distributed to unauthorized people outside of the course. Any suspected instances of a breach of academic integrity will be reported to the Academic Integrity Office for review.

LATE ASSIGNMENTS AND QUIZZES Late assignments will be subject to a 10% deduction per day (note that assignments handed in after the first 5 minutes of lab are considered late) up to a maximum of 2 days late (after which you will receive a 0). There are no make-up quizzes offered except in the case of a documented medical or family emergency (in which case the instructor will decided how to go about the make-up testing). No late participation items will be accepted as only up to 80% of these grades are counted.

LABORATORY ATTENDANCE – Enrolled and waitlisted students MUST attend the first lab session. Additional details: http://biology.ucsd.edu/go/ug-labs.

Attendance in laboratory is required. Missing one laboratory session without a documented excuse (documented illness or serious family emergency), will automatically result in a 5% deduction in your final course grade. If you miss two labs for undocumented reasons, you will be asked to drop the course. Please be on time for laboratory sessions. Two late attendances will be counted as one absence. Additional policies are available online (https://biology.ucsd.edu/education/undergrad/course/waitlist.html).

ADD/DROP DEADLINES are different for lab courses than lecture courses. <u>Students who drop a Biology lab class after the end of the second class meeting will be assigned a "W"</u>. Additional details: <a href="http://biology.ucsd.edu/go/ug-labs">http://biology.ucsd.edu/go/ug-labs</a>.

LAB SAFETY TRAINING – Enrolled and waitlisted students MUST successfully complete the Biology Lab Safety Training and Assessment before the first lab session: <a href="https://dbsportal3.ucsd.edu:3443/safety-training/">https://dbsportal3.ucsd.edu:3443/safety-training/</a>. Please note that courses offered by other departments (Chemistry, for example) may have additional safety training requirements.

If a student arrives at the <u>first lab session</u> having not passed the safety assessment they may be allowed to complete the lab at the discretion of the instructor if the planned activities and environment do not pose any lab safety hazards. Students are not allowed into the lab for the second lab session unless they have successfully passed the safety assessment.

#### WRITING CENTER -- https://writingcenter.ucsd.edu/

The Writing Center provides support for undergraduates working on course papers (i.e. laboratory reports and the research proposal) and independent writing projects. Writing mentors can help at any stage of the writing process, from brainstorming to final polishing. The Writing Center offers: one-on-one appointments for undergraduates with peer writing mentors; group workshops addressing a variety of writing projects, genres, and issues; and Drop-In Zone for quick questions, targeted assistance, and a comfortable writing space.

TECHNOLOGY POLICY: Laptop computer policy: Students are welcome to bring laptops to lecture for note-taking purposes. Please see this research study that shows "multi-tasking" on computers is likely to decrease your grade, but it also decreases the grades of people around you who can see your screen<sup>2</sup>! For this reason, we ask that you do not flip between lectures notes and the internet. The use of cell phones, computers, or any other electronic devices is not permitted during exams. Use of a cell phone or other similar electronic devices during an exam or quiz is grounds for receiving a failing grade.

2 Sana et al. 2013. http://www.sciencedirect.com/science/article/pii/S0360131512002254

/eek	Date	Class or Lab	Section of Lab Manual	Quiz/Report Due Date
	9/28/18	Course Introduction, Lecture 1		-
0		LAB 1		
	0/29/19	Calibration of a pipettemen	Lab 1	
	9/28/18	Dinetting	Additional info "working in the lab" sections D, E, F, G	
		Pipetting Dilutions	Sections D, E, F, G	
	10/1/18	Lecture 2		
	10/3/18	Lecture 3		<b>-</b>
		LAB 2		
	10/3/18	Agarose gel electrophoresis on two DNA samples of		
		unknown size and concentration (estimating using standard curve)	Evporiment 1 1 A 1 D	
1	10/5/18	Lecture 4	Experiment 1, 1A-1D	
	10/5/16	LAB 3		1
	10/5/18	Computer Lab		
		Image Studio Lite Analysis of Agarose Gel	Appendix A	
	10/0/10	Graphing	Appendix B, C	
		Set-up liquid cultures of RFP and control promoter	Starting Experiment 2, 2A	
	10/8/18			Quiz 1
	10/10/18	LAB 4		1
		Extract plaemide		
	10/10/18	Check plasmids with AGE & nanodrop	2B	
2		Computer Labs available if needed	20	
2	10/12/18	Lecture 6		
	10,12,10	LAB 5		
		Design and set up RFP PCR experiment	Sub-experiment 2-1, 2C	
	10/12/18	Start computer lab - plasmid map, restriction enzymes,	·	
		designing primers	Appendix D	
		Lecture 7		
	10/17/18	Lecture 8		•
		LAB 6	F: 11.00	
	10/17/18	Run gel of PCRs, repeat if needed	Finish 2C	Mini Report 1 due in L
		Set up digest of Pro1 plasmid and RFP PCR product	2D 2E	
3		Finish Appendix D computer lab	2L	
	10/19/18	Lecture 9		_
	10,10,10	LAB 6		1
	40/40/40	Clean stuffer from Pro1 - heat inactivate PCR digest	2F	
	10/19/18	Run gel of digest	2F	
		Plan LIGATIONS	Sub-experiment 2-2: part of 2G	
	10/00/10			
	10/22/18	Lecture 10		Quiz 2
	10/24/16	LAB 8		1
	10/24/18	Set-up ligations & transform bacteria with ligations	2H	
	10/24/10	Computer Lab: Design mutagenesis primers	2K	
4	10/26/18	Lecture 11		
		LAB 9		1
	10/26/18	Count colonies	21	
	10/20/10	Plan how to analyze ligation data	start 2I	
		Pick red colony from plate and start liquid culture	21	
	10/00/40	Leature 10		
		Lecture 12		
	10/31/18	Lecture 13		
		Purify recombinant Pro1 PEP placmid and run gol	31	Mini Donort 2 duo in L
	10/31/18	Purify recombinant Pro1-RFP plasmid and run gel Set up mutagenesis PCR	2J 2L	Mini Report 2 due in L
		Computer lab: analyze ligation data	plan previously developed	
5		Timp side take analyze ngadon data	p.a promodely dovoloped	_
5	11/2/18	Lecture 14		
5	11/2/18	Lecture 14		
5		LAB 11	2M	
5	11/2/18		2M 2N	

Week	Date	Class or Lab	Section of Lab Manual	Quiz/Report Due Dates
	11/5/18			Quiz 3
	11/7/18	Lecture 15		-
		LAB 12		
6	11/7/18	Check repeat PCRs, KLD and transformation if needed	20	
		Analyze transformations Computer lab: Bioinformatics Intro to GenBank (optional)	20 Appendix F	
	11/9/18	Lecture 16	Appendix i	
		LAB 13		1
	11/9/18	Set-up liquid cultures: three colonies from mutagenesis	20	
		Holiday - no lecture		
	11/14/18	Lecture 17		=
		LAB 14		
	11/14/18	Streak cultures to maintain	2P	Mini Report 3 due in Lab
		Purify plasmids from 3 cultures and send for sequencing	2Q 2Q	
7	11/16/19	Check plasmids using AGE Lecture 18	ZQ	
	11/10/10	LAB 15		
		Computer lab: analyze sequencing results	2R	
	11/16/18	Use streaked bacteria to measure RFP	2S	
		Plan how to analyze RFP data (optional: analysis as		
		homework or anlayze next lab)	start 2T	
	11/19/18			Quiz 4
	11/21/18	Lecture 19		-
8	11/21/18	LAB 16	F : 10.04	
		Observe C.elegans and induce RNAi Computer Lab: Analzye RFP data	Experiment 3. 3A 2T	
	11/23/18	Thanksgiving Holiday - no lecture or lab		
	11/20/10	a.mogga.aya.acta.a oa.a		
		Lecture 20		
	11/28/18	Lecture 21		_
	11/28/18	LAB 17		Mini Report 4 due in Lab
		Observe worm phenotypes	3B	
9	44/20/40	Extract RNA and set up RT-qPCR Lecture 22	3C	
	11/30/18	LAB 18		1
	11/30/18	PTC extraction & PCR	Experiment 4. 4A	
		Analyze qPCR data *need own computers, only laptops in	Experiment 4. 47	
		3rd floors labs available	Brief instructions at end of Exp. 3.	
	12/3/18	Lecture 23		
	12/5/18			_
	12/5/18	LAB 19		
				RNAi/PTC
		Digest PTC PCRs, check with agarose gel, PTC taste-test	15	assignments due
10		(phenotyping)	4B	online
		Pool genotype/phenotype data Analyze data	4B	
	12/7/18	Final class meetingCoffee and Donuts		
	,,,,	LAB 20		
	12/7/18	Clean-up & final quiz		Final Quiz
		Computer labs available		